Vetscan Imagyst® Digital Cytology

THE BASICS

Staining and Submission

Keys to Excellent Stain Quality

- Always stain samples using a quick stain (e.g., Diff Quik®, quick aqueous or other Romanowsky-type stain) according to manufacturer's protocol
- A dip method is preferred to flooding the slides with stain to ensure uniform staining
- Stains should be replenished regularly to avoid depletion and build up of stain precipitate
- Avoid heating, freezing, or refrigerating cytology slides, which may distort cells
- Ensure unstained cytology slides are not collected in the vicinity of formalin or formalin vapors. Exposure of cytology specimens to formalin prior to staining may make the slides uninterpretable as formalin interferes with cell staining
- **Alternative staining techniques such as Gram staining or urine sediment (supravital) stain are not acceptable for submission**

Coverslipping

- Use Zoetis issued or similar coverslip (24 x 60 mm; 0.13-0.17 mm thickness)
- Ensure that only one coverslip is used as coverslips can easily stick together
- Always use a coverslip

1. Place stained slide on flat surface.
2. Add 2 drops of immersion oil to surface of sample. Use only enough to cover sample. Excess oil can contaminate scanner lens.
3. Handle coverslip by its edge to avoid fingerprints.
4. Place edge of coverslip onto sample and roll over sample, avoiding formation of air bubbles.
5. Blot excess oil with a delicate task wipe or lotion free tissue.

Scanning

Slide

- Place sample side up, label to the right
- Flat on stage, stage clips flush with edges of the slide
- Slide lock engaged

Mask

- Masked area(s) completely within bounds of coverslip
- Avoid masking large areas that do not contain much stained material

Scanner

- Lens and stage are clean
- No objects nearby scanner that may impede movement
- No nearby centrifuges in operation during scan